

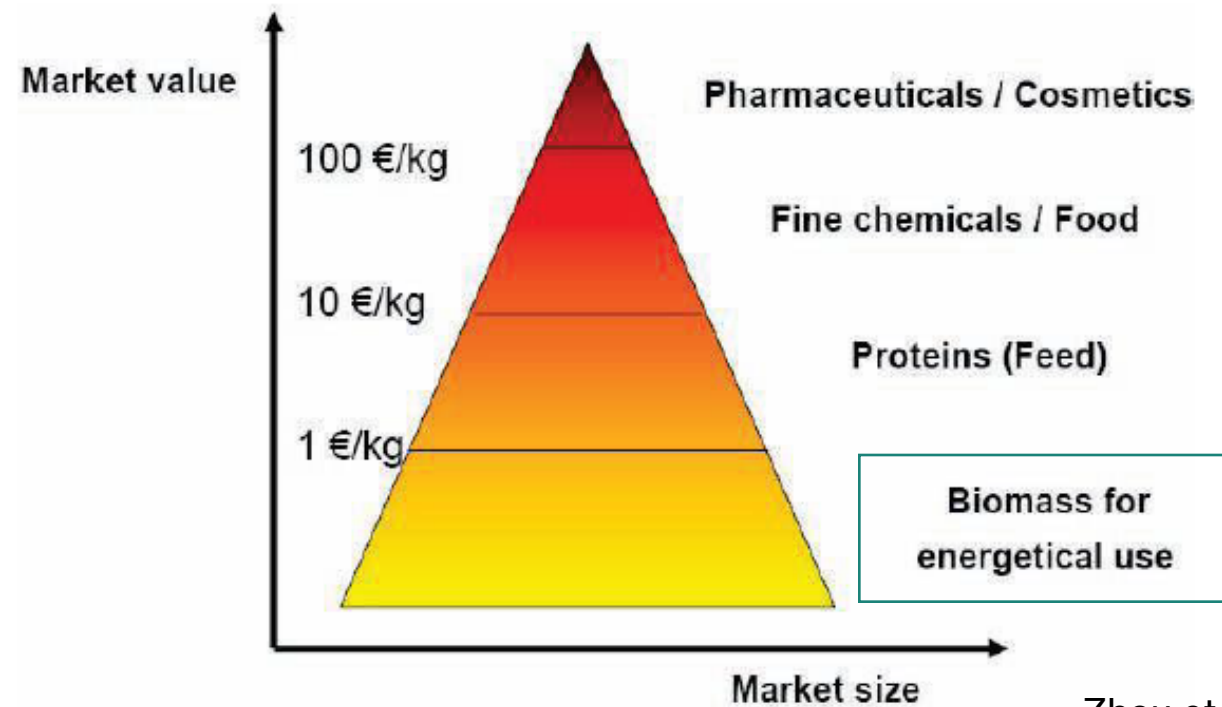


Microalgae

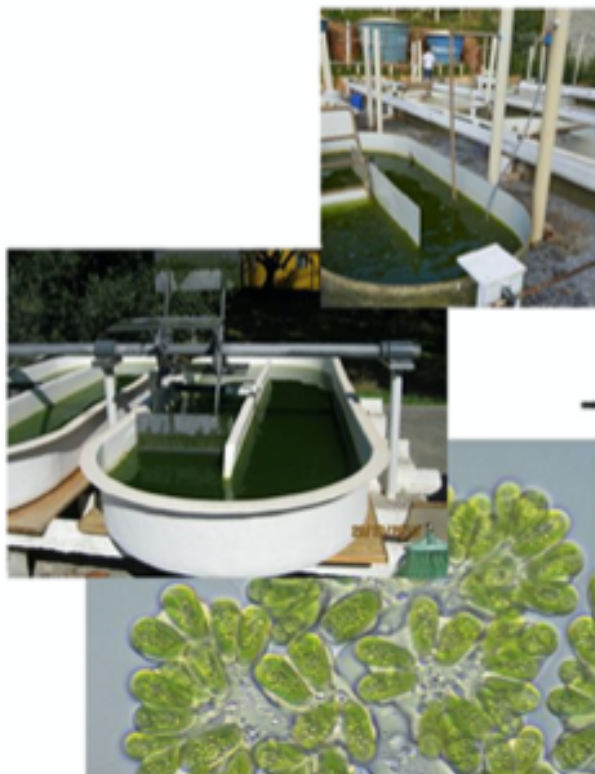
Energy valorization routes

Wastewater grown microalgae biomass for energy purposes

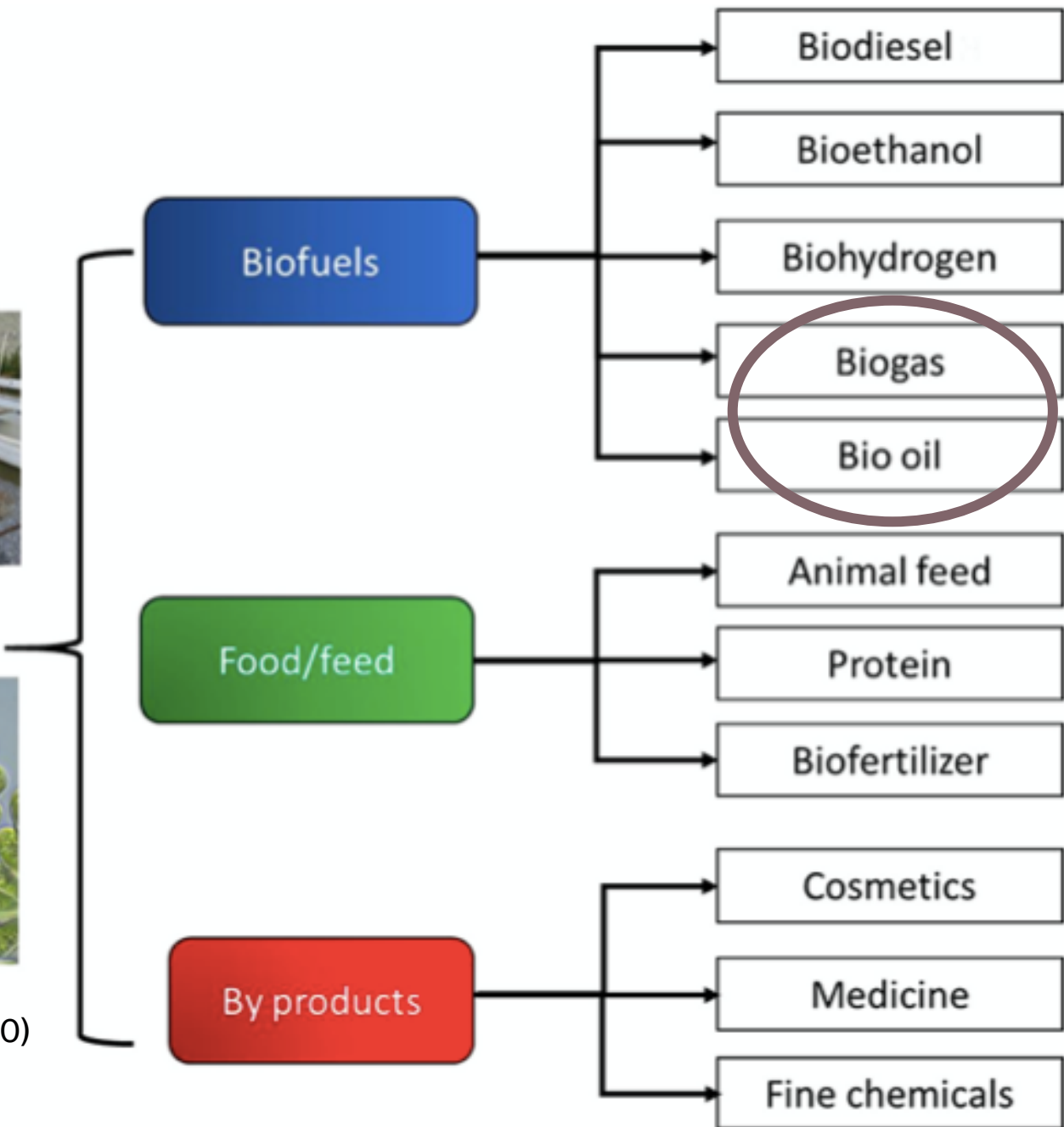
- Biomass composition as a function of specie, growth conditions...
- Low lipid content
- Less noble uses



Zhou et al. (2015)



Couto et al. (2020)



Microalgae anaerobic digestion - challenges

Biomass	Methane (m ³ /kg VS)
Solid waste	0.2-0.53
Vegetables waste	0.42
Microalgae	0.26
Wood waste	0.20
Corn waste	0.31

Kwietniewska and Tys (2014)



Low C/N ratio: co-digestion



Cell wall: pre-treatment

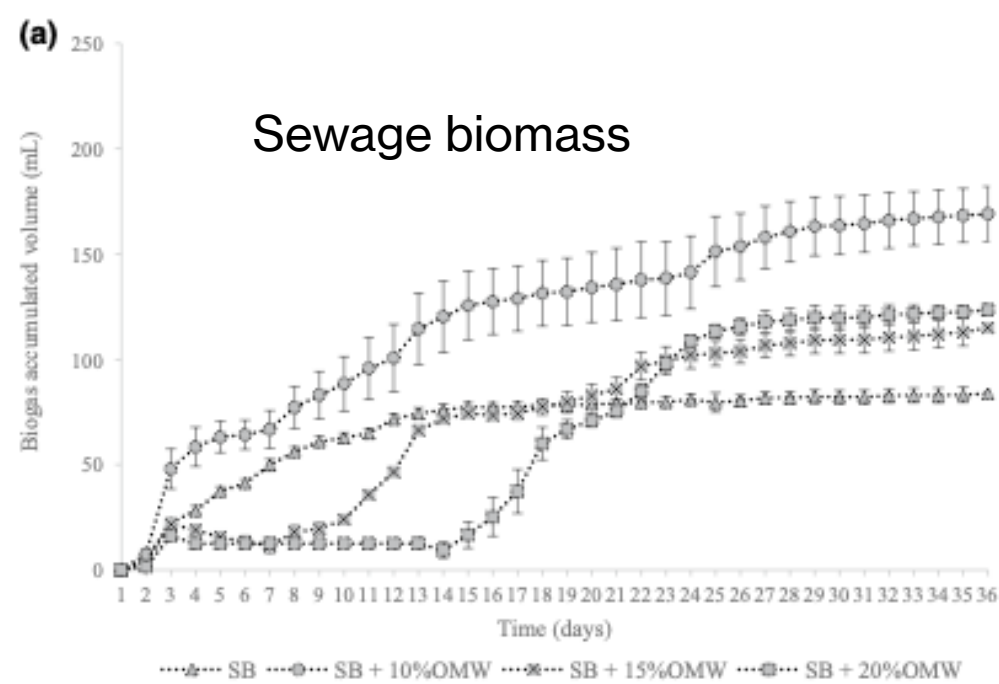
Table 4

Effect of physical and chemical pre-treatment of wastewater grown microalgae on methane yield during anaerobic digestion (AD).

Pre-treatment method	Operational conditions	Microalgae, cultivation medium	Methane yield (m ³ CH ₄ /kg VS)	Yield improvement	References
Physical, ultrasound	67 MJ/kg TS, 20 kHz	<i>Scenedesmus</i> , <i>Chlorella</i> , domestic sewage	0.169	33%	Passos et al., 2014a
Physical, microwave	110,500 kJ/kg VS, 900 W (output power), 3 min (exposure time)	<i>Monoraphidium</i> sp., <i>Stigeoclonium</i> sp., <i>Scenedesmus</i> sp., <i>Nitzschia</i> sp. domestic sewage	0.20	60%	Passos et al., 2014c
Physical, microwave	2450 MHz, 700 W (output power), 840 s (exposure time)	Microalgal bacterial flocs, pikeperch aquaculture wastewater	0.19	7%	Van Den Hende et al., 2015
Chemical, acid	Addition of HNO ₃ to pH 6.3	Microalgal bacterial flocs, Pikeperch aquaculture wastewater	0.17	34%	Van Den Hende et al., 2015
Chemical, lipid extraction	Addition of H ₂ SO ₄ + lipid extraction	<i>Scenedesmus</i> sp., meat processing industry wastewater	2.4 ^a	5 times	Assemany et al., 2016
Chemical, lipid extraction	Addition of H ₂ SO ₄ + lipid extraction	<i>Scenedesmus</i> sp. <i>Chlorella vulgaris</i> , domestic sewage	2.6 ^a	10 times	Assemany et al., 2018a, 2018b
Chemical, alkaline	Addition of 4% CaO	<i>Chlorella</i> sp. and <i>Scenedesmus</i> sp., synthetic wastewater	0.282	19.8%	Solé-Bundó et al., 2017a
Chemical, alkaline	Addition of 10% CaO	<i>Chlorella</i> sp. and <i>Scenedesmus</i> sp., synthetic wastewater	0.259	14.9%	Solé-Bundó et al., 2017a
Chemical, thermo-alkaline	Addition of 10% CaO + 72 °C	<i>Chlorella</i> sp., <i>Monoraphidium</i> sp. and diatoms, domestic sewage	0.287	9%	Solé-Bundó et al., 2017b

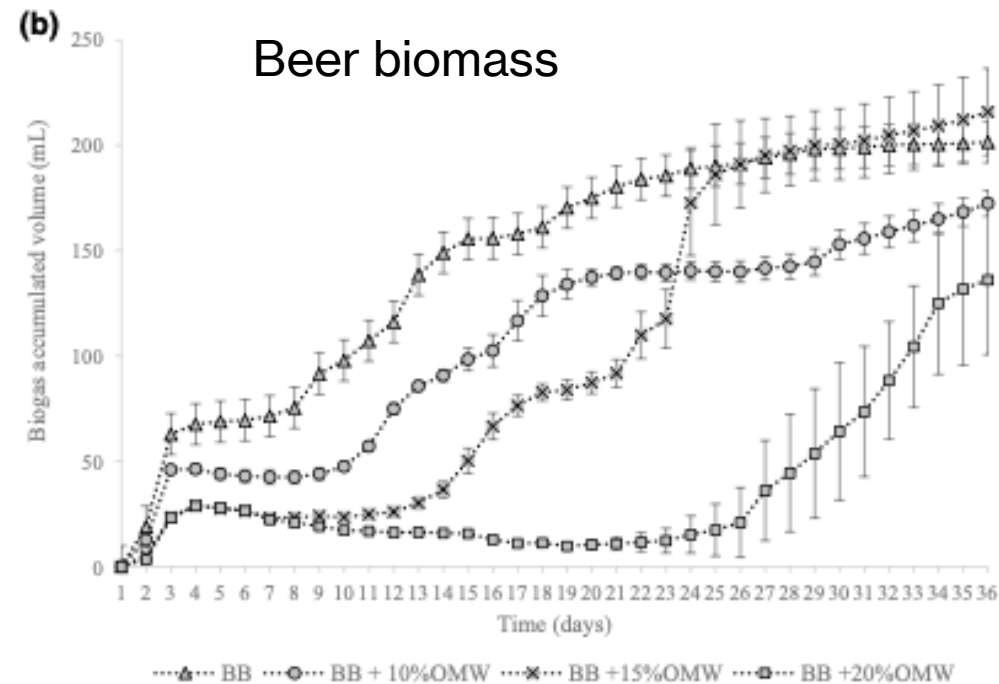
^a Biogas yield.

Choudhary et al. (2020)



SB+10%OMW: 0.10 m³ CH₄/kg VS

SB: 0.06 m³ CH₄/kg VS



BB: 0.16 m³ CH₄/kg VS

BB +10% OMW: 0.13 m³ CH₄/kg VS

Anaerobic digestion: next steps

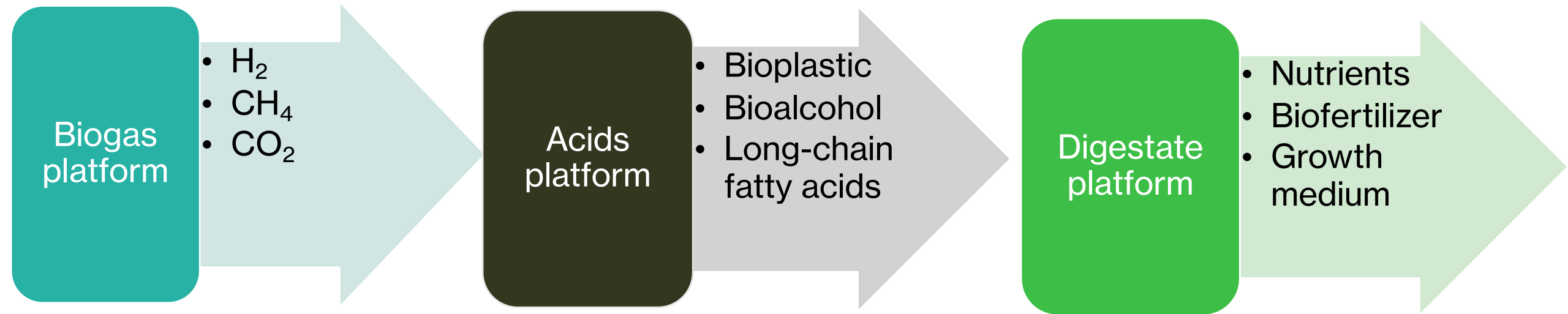
Less energy demand pre-treatment methods

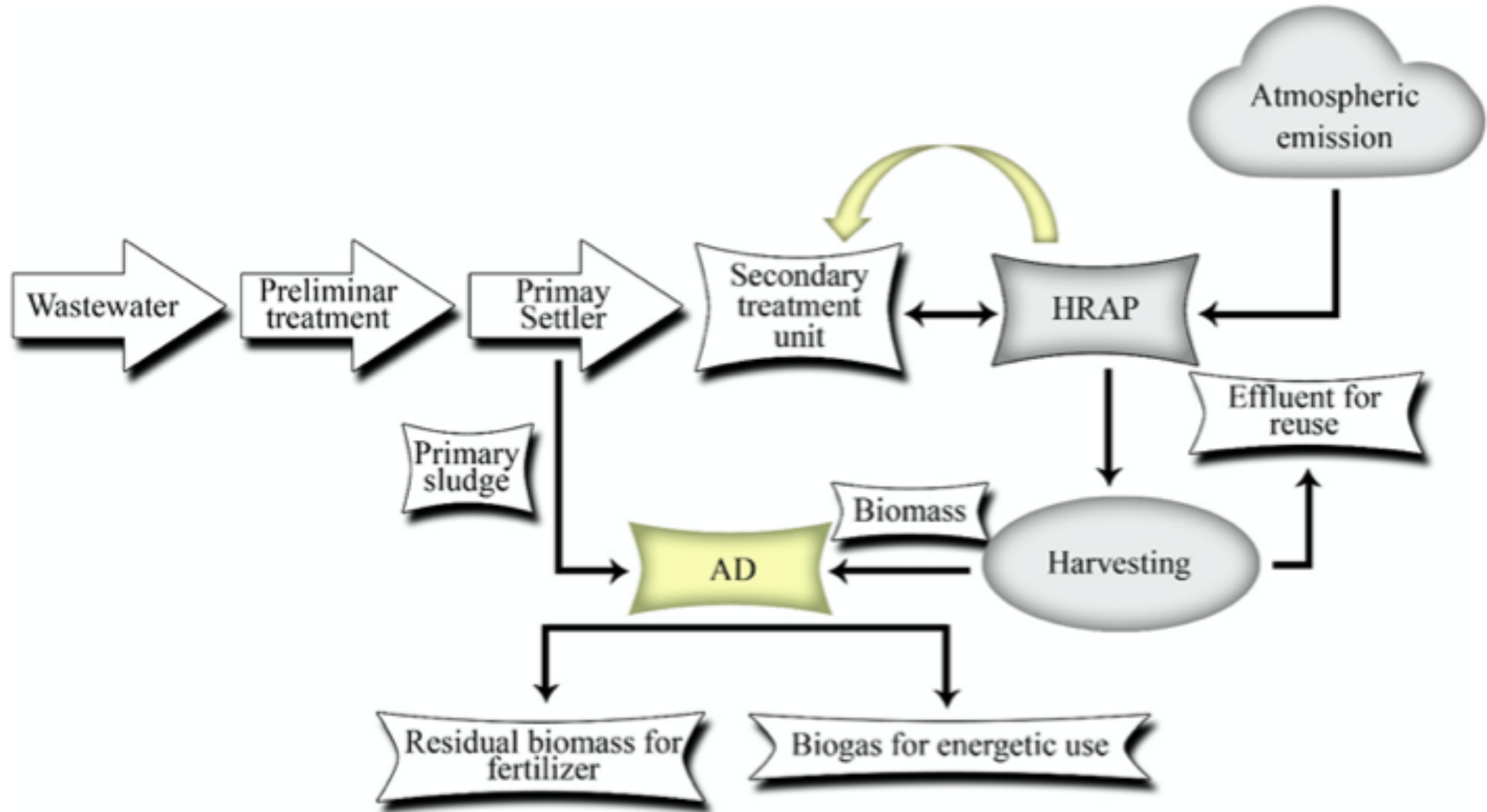
Reactors designed for algal biomass digestion

Full scale

Anaerobic digestion as a biorefinery
– new routes, new by-products

Anaerobic digestion as an industrial process





Choudhary et al. (2020)

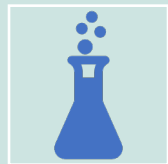
Hydrothermal carbonization and liquefaction



High ash content



N in the bio-oil



Catalysts



Subproducts
valorization

Table 1

Comparative performance of different catalysts for the production of biocrude obtained from HTL of algal biomass grown under varied nutrient medium.

Catalyst	Reaction conditions	Microalgae	Nutrient medium/wastewater	Biocrude yield (%) & HHV (MJ/kg)	Reference
K ₂ CO ₃	T= 200 (°C), P= 2.5 (MPa N ₂), t= 20 (min)	A consortium of <i>Chlorella</i> & <i>Phormidium</i> sp.	Artificial medium	42.0 & 33.10	Choudhary et al., 2020
Cerium oxide (CeO ₂)	T= 250 (°C) ; t= 30 (min)	<i>Chlorella</i> & <i>Phormidium</i>	Wastewater		Naaz et al., 2019
CaO	T= 320 (°C) ; t= 10 (min)	<i>Spirulina Platensis</i>	Artificial medium	26.0 & 27.15	Kandasamy et al., 2020
Pt/C	T= 350 (°C), P=0.3 (MPa H ₂), t= 30 (min)	<i>Nannochloropsis gaditana</i>	Commercial source	49.73 & 31.47	Sánchez-Bayo et al., 2020
ZrO ₂ /SO ₄ ²⁻ , HZSM-5, MgO/MCM-41	T= 350 (°C); t= 60 (min)	<i>Chlorella</i> sp.	Commercial source	37.09 & 29.73	Xu et al., 2019
KtB		<i>Dunaliella tertiolecta</i>	Commercial source	28.0 & 33.24 30.0 & 33.67 35.0 & 33.17 49.0 & 32.36	Chen et al., 2015
K ₂ CO ₃	T=260-320 (°C); t= 10-40 (min)	<i>Chlorella vulgaris</i>	Anaerobic digestion effluent (ADE)	41.2	(Liang et al., 2015)
HZSM-5		<i>Spirulina</i>	grown (5% conc.)	27.7	
		<i>Raceway biomass</i>		31.4	
Ru/C	T= 320-400 (°C); t= 20 (min)	<i>Nannochloropsis</i> sp.	Commercial source	45.0 & 38.92	(Xu and Savage, 2015)
Catalyst	Reaction conditions	Microalgae	Nutrient medium/wastewater	Biocrude yield (%) & HHV (MJ/kg)	Reference
Co/Mo/Al ₂ O ₃	T= 350 (°C); t= 60 (min)	<i>Chlorella vulgaris</i>	Effluent from wastewater treatment plant (WWTP)	38.7 & 39.7	(Roberts et al., 2013)
Ni/Al/Al ₂ O ₃		<i>Nannochloropsis occulta</i>		30.0 & 42.0	
Pt/Al/Al ₂ O ₃				38.9 & 38.2	
Non-catalytic	T=325 (°C), t=45 (min); P= 12 (MPa)	<i>Monoraphidium</i> sp. + domestic sewage sludge (DSS)	Sewage treatment plant effluent	39.38 & 39.47	Mishra and Mohanty, 2020
Non-catalytic	T= 260-320(°C), t=45 (min); P= 12 (MPa)	<i>Nannochloropsis</i> sp. and <i>Sargassum</i> sp.,	Fish Park aquarium	39.05-54.11 & 35.92-37.88	He et al., 2020
Non-catalytic	T= 325-350 (°C), P=20 MPa; t= 3-9 (min)	<i>Galdieria sulphuraria</i> (monoculture & polyculture)	Municipal wastewater	3.11-9.49 & 33.63-35.23	
Non-catalytic	T= 275-350 (°C), t= 30 (min); P= 0.6-0.8 (MPa)	<i>Tetraselmis</i>	Sea-water	28.1 & 39	Cheng et al., 2019
Non-catalytic	T= 300 (°C); P= 9 MPa; t= 30 (min)	<i>Kirchneriella</i> sp.	Commercial source	26.3-31.0 & 29.5-33.3	Han et al., 2019
Non-catalytic	T= 150-300 (°C); t= 60 (min)	Algal consortia (<i>Pediastrum</i> sp., <i>Micractinium</i> sp., and <i>Desmodesmus</i> sp.)	Effluent from WWT HRAPs	45.5 & 37.52	Dandamudi et al., 2019
				24.89 & 38.9	(Mehrabadi et al., 2017)

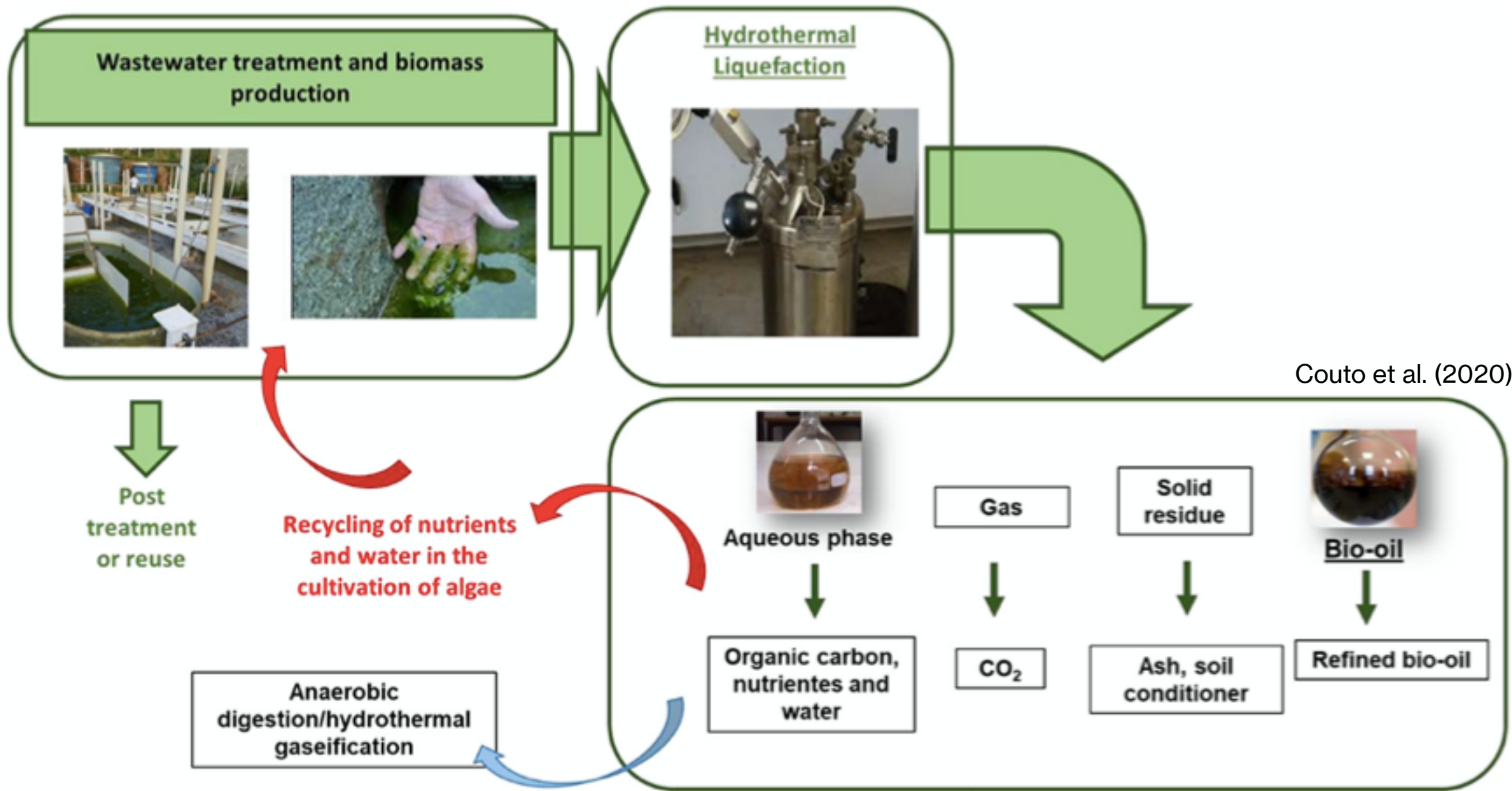
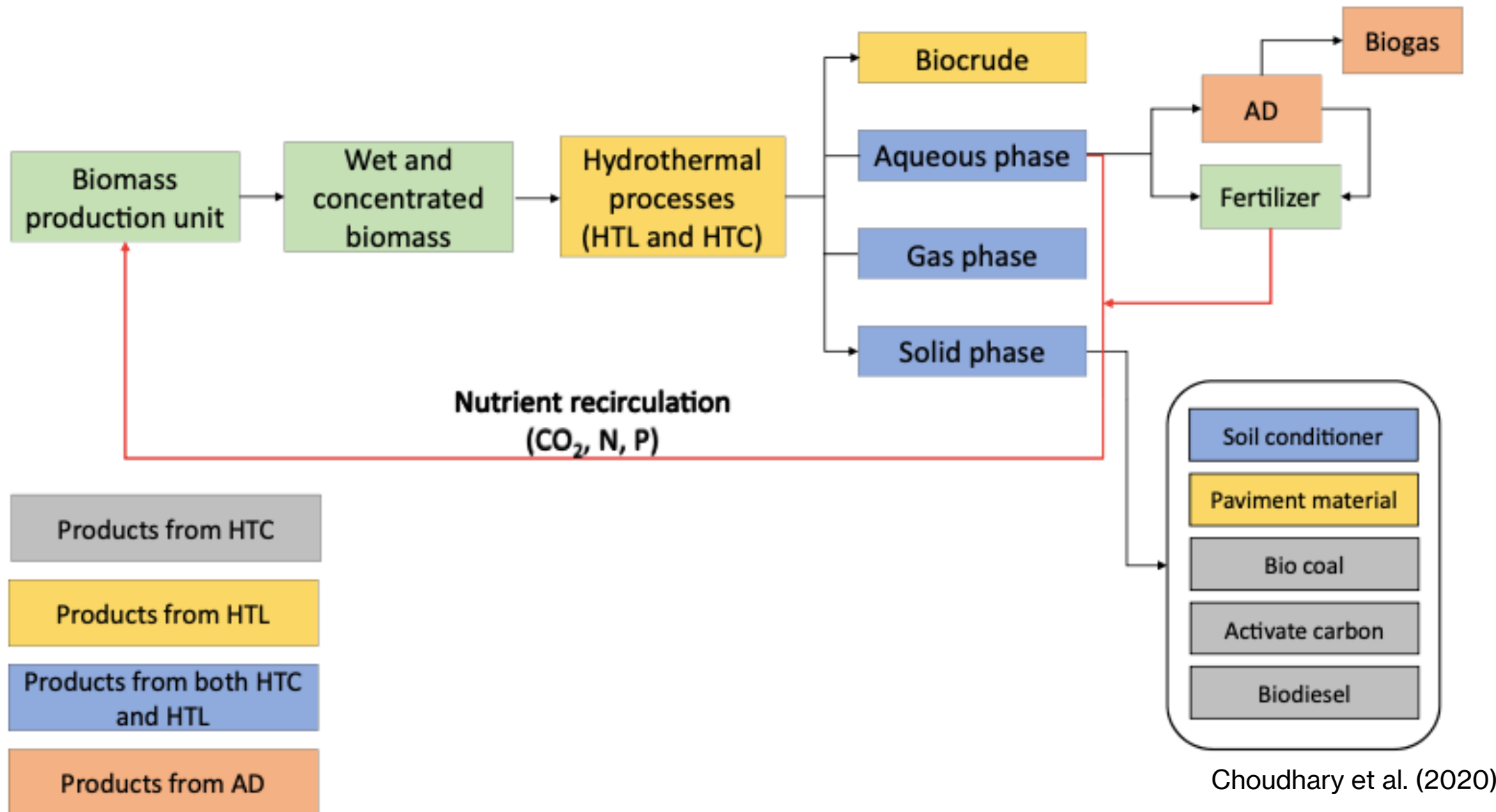


Table 2

Studies on hydrothermal carbonization (HTC) of synthetic media grown algal biomass.

Biomass	Reaction conditions	Catalyst	HHV (MJ/kg)	Yield (%)	References
<i>Aphanizomenon flos-aquae</i> – char	213 °C; 120 min	Citric acid	–	16.0	Heilmann et al., 2010
<i>Synechocystis</i> sp. – char	213 °C; 120 min	Citric acid	–	18.0	
<i>Spirulina</i> sp. – char	213 °C; 180 min	Oxalic acid	–	27.0	
<i>Chlorella</i> sp. – char	200 °C; 180 min	Oxalic acid	–	39.5	
<i>Dunaliella salina</i> – char	200 °C; 180 min	Oxalic or acid citric	30.5	36.0	Levine et al., 2013
<i>Chlamydomonas reinhardtii</i> – raw biomass	–	–	18.0	–	
<i>Chlamydomonas reinhardtii</i> – char	200 °C; 120 min	Oxalic acid	31.6	39.0	
<i>Nannochloropsis oculata</i> – raw biomass	–	–	20.6	–	
<i>Nannochloropsis oculata</i> – char	180 °C; 15 min	–	26.3	51.0	Levine et al., 2013
<i>Nannochloropsis oculata</i> – char	180 °C; 30 min	–	26.9	49.0	
<i>Nannochloropsis oculata</i> – char	190 °C; 15 min	–	26.8	51.0	
<i>Nannochloropsis oculata</i> – char	190 °C; 30 min	–	27.1	47.0	
<i>Nannochloropsis oculata</i> – char	200 °C; 15 min	–	27.3	47.0	Bach et al., 2013
<i>Nannochloropsis oculata</i> – char	200 °C; 30 min	–	27.3	44.0	
<i>Nannochloropsis oculata</i> – char	210 °C; 15 min	–	28.0	45.0	
<i>Nannochloropsis oculata</i> – char	210 °C; 30 min	–	28.4	41.0	
<i>Chlorella vulgaris</i> – raw biomass	–	–	22.0 ^{daf}	–	Ekpo et al., 2016
<i>Chlorella vulgaris</i> – char	160 °C; 10 min	–	23.8 ^{daf}	61.7	
<i>Chlorella vulgaris</i> – char	170 °C; 10 min	–	24.2 ^{daf}	~59.0	
<i>Chlorella vulgaris</i> – char	180 °C; 10 min	–	24.5 ^{daf}	52.6	
<i>Chlorella vulgaris</i> – char	170 °C; 5 min	–	23.6 ^{daf}	62.9	Lee et al., 2018
<i>Chlorella vulgaris</i> – char	170 °C; 30 min	–	26.0 ^{daf}	51.8	
<i>Chlorella vulgaris</i> – raw biomass	–	–	21.1	–	
<i>Chlorella vulgaris</i> – char	250 °C; 60 min	–	14.4	~15.0	
Lipid extracted <i>Chlorella vulgaris</i> – raw biomass	–	–	–	–	Lee et al., 2018
Lipid extracted <i>Chlorella vulgaris</i> – char	180 °C; 30 min	–	~25.00	74.53	
Lipid extracted <i>Chlorella vulgaris</i> – char	200 °C; 30 min	–	~27.00	72.56	
Lipid extracted <i>Chlorella vulgaris</i> – char	240 °C; 30 min	–	~23.00	51.84	
<i>Spirulina platensis</i> – raw biomass	190 °C; 60 min	–	21.64	–	Zhao et al., 2019
<i>Spirulina platensis</i> – char	190 °C; 60 min	–	29.63	~30.00	
<i>Chlorella vulgaris</i> – raw biomass	190 °C; 60 min	–	27.00	–	
<i>Chlorella vulgaris</i> – char	190 °C; 60 min	–	20.79	~37.00	

^{daf} Dry-ash-free-basis; – indicates that the data was not available.



Choudhary et al. (2020)

Paula Assemany

Environmental Engineering Department
Engineering School
Federal University of Lavras, Brazil

paula.assemany@ufla.br
paula_assemany@hotmail.com



Thank you!